

The effects of selenium and conjugated linoleic acid (CLA) isomers on fatty acid composition, CLA isomer content in tissues, and growth of rats*

**M. Czauderna¹ J. Kowalczyk, I. Wąsowska, K. M. Niedźwiedzka
and B. Pastuszewska**

*The Kielanowski Institute of Animal Physiology and Nutrition,
Polish Academy of Sciences
05-110 Jabłonna, Poland*

(Received 21 July 2003; accepted 28 October 2003)

ABSTRACT

The effects of conjugated linoleic acid (CLA) isomers and/or selenium (as Na₂SeO₄) on rat growth performance and levels of some fatty acids (FA) in femoral muscles and perigonadal fat were investigated. Feeding a mixture of CLA isomers, the *trans*-10,*cis*-12 isomer, or selenium (Se) tended to decrease the rats' body weight compared with the control animals, whereas a diet supplemented with both Se and the *trans*-10,*cis*-12 isomer produced the highest increase in body weight gain and the highest feed conversion efficiency. None of the experimental diets had any influence on brain, kidney, pancreas, or liver weight, while they significantly increased spleen mass. The administered CLA isomers significantly increased the contents of CLA isomers and non-CLA fatty acids containing conjugated double bonds (CD) in the assayed tissues. Se and individual CLA isomers or their mixture increased the CLA isomer and CD contents in muscles, while feeding only the *cis*-9, *trans*-11 isomer and Se resulted in an increase in CD and sum of all CLA isomers in fat. These results demonstrate that the *trans*-10,*cis*-12 isomer is preferentially driven through the β-oxidation pathway in muscles and fat compared with the *cis*-9,*trans*-11 isomer. CLA isomers added to Se-unsupplemented or -supplemented diets increased the levels of total FA in muscles only, while decreased the level of total assayed saturated FA in fat. Feeding Se and individual CLAs or a mixture of these isomers resulted in increasing both the sum of *cis*-monounsaturated FA (*cis*-MUFA), CD and polyunsaturated FA (PUFA) in muscles, but generally resulted in decreasing the level of total saturated FA, *cis*-MUFA and PUFA in fat. The interaction between Se and a mixture of CLA isomers or the *trans*-10,*cis*-12 isomer most effectively protected CLA isomers from peroxidation damage and/or catabolism in muscles.

KEY WORDS: rat, CLA, isomers, conjugated dienes, selenium, body gain, feed intake

* Supported in part by the State Committee for Scientific Research, Grant No 3 PO6Z 034 22

¹ Corresponding author: e-mail: r.czauderna@ifzz.pl

INTRODUCTION

Conjugated linoleic acid (CLA) isomers (predominant isoforms: *cis-9,trans-11* and *trans-10,cis-12*), which are mainly present in ruminant meat and milk fat (Jahreis et al., 1997; Bauman et al., 2000), are believed to play a principal role in many important physiological functions and may act to convey body composition or to modulate immune function. In ruminants or monogastric animals, the optimal content of CLA isomers is highly correlated with adiposity and overall energy expenditure. CLA isomers are also believed to inhibit tumorigenesis by interfering in the metabolism of some carcinogens. The specific CLA isomers responsible for these biological effects have not been clearly established, although the physiologically most active isomer, *cis-9, trans-11*, is thought to play a key role in the effect of CLAs on atherosclerosis and in their anticarcinogenic properties. Moreover, many authors found some beneficial health effects of another CLA isomer, *trans-10,cis-12*, on lipid metabolism such as body fat reduction with enhancement of lean body mass (Sébédio et al., 1999; Alasnier et al., 2002). Several studies give support not only to CLAs as effective antioxidants, but also to their ability to increase fatty acid oxidation. An increased hydroxybutyrate:acetoacetate ratio has led some authors to postulate that the dietary *trans-10,cis-12* isomer appears to exert its hypolipidemic effect by increasing β -oxidation of fatty acids at the expense of fatty acid esterification. In agreement with these results, many other recent studies have demonstrated that feeding CLAs enriched in the *trans-10,cis-12* isomer is a more potent antiobesity agent in rodents compared with other CLA isomers. Several recent studies had reported an inverse relationship between CLA and blood plasma leptin in rats and mice (DeLany et al., 1999; Tsuboyama-Kasaoka et al., 2000; Rahman et al., 2001). Moreover, the *trans-10, cis-12* isomer reduced excess weight gain, lowered blood lipids and improved feed efficiency. Early studies using rats found that the content of polyunsaturated fatty acids (PUFA), especially in serum cholesterol esters and phospholipids, was positively correlated with the selenium (Se) concentration in the diet (Crespo et al., 1995). Se, an essential nutritional trace element for mammals, has been found to be an integral part of the active site of cytosolic and mitochondrial glutathione peroxidases as well as phospholipid hydroperoxide glutathione peroxidase (cGPx) (Wolffram, 1999; Zagrodzki et al., 2000, 2001; Arteel and Sies, 2001). Furthermore, studies have established that phospholipid hydroperoxide cGPx acts more directly in protecting PUFA from peroxidation damage (Crespo et al., 1995).

The potential importance of CLA isomers in living organisms, as well as that of Se, essentially related to protection against oxidative stress, made it desirable to study the extent to which dietary CLA isomers may contribute to the CLA isomer levels in femoral rat muscle and perigonadal fat. The objective of the present experiment was also to investigate the effect of dietary Se (as sodium selenate) on

some other fatty acid concentrations in these rat tissues. Moreover, the influence of the experimental factors on the body gain of rats, organ weight, and feed intake was also determined.

MATERIAL AND METHODS

Animals and diets

Ten groups of female rats (Wistar, Ifz: BOA), at 8 weeks of age, each weighing about 200 g at the beginning of the experiment, were housed individually in plastic cages at a constant temperature ($22 \pm 1^\circ\text{C}$) with 12 h light-dark cycle and relative humidity 50-60%. Each group comprised 7-8 animals. During a 1-week preliminary period the animals were fed a standard Labofeed diet produced by the Feeds and Concentrates Production Plant in Kcynia, A. Morawski (Poland) (Pastuszewska et al., 2000) given at a submaintenance level to reduce body fat content. During that time they decreased body weight by about 15 g per animal. Then they were fed the experimental diets (Table 1) enriched with a CLA isomer mixture, *cis*-9,*trans*-11 isomer (*c9,t11*), *trans*-10,*cis*-12 (*t10,c12*) isomer or/and Se (as Na_2SeO_4). The rations were adjusted each day to ensure the *ad libitum* feeding level. Feed intake and body weight of rats were measured weekly. No lesions or symptoms of Se or CLA isomer intoxication were found in rats fed the experimental diets. After the four-week experimental period the rats were killed

TABLE 1

Diet composition

Group	Supplement	Concentration
1 (control)	-	-
2 _{+Se}	selenium	2 ppm
3	CLA isomer mixture ¹	1 %
4	<i>cis</i> -9, <i>trans</i> -11 isomer (<i>c9,t10</i>)	1 %
5	<i>trans</i> -10, <i>cis</i> -12 isomer	1 %
6	CLA isomer mixture	2 %
3 _{+Se}	selenium	2 ppm
	CLA isomer mixture	1 %
4 _{+Se}	selenium	2 ppm
	<i>cis</i> -9, <i>trans</i> -11 isomer (<i>c9,t11</i>)	1 %
5 _{+Se}	selenium	2 ppm
	<i>trans</i> -10, <i>cis</i> -12 isomer (<i>t10,c12</i>)	1 %
6 _{+Se}	selenium	2 ppm
	CLA isomer mixture	2 %

¹ concentration ratio of the *cis*-9,*trans*-11 isomer to the *trans*-10,*cis*-12 isomer in dosed CLA standard: 1.0242 (Czauderna et al., 2003)

and the femoral muscles and perigonadal fat samples were quickly removed and weighed.

Chemicals

All reagents were of analytical grade, whereas organic solvents were HPLC grade. Acetonitrile and n-hexane (95%) were purchased from Lab-Scan (Ireland), while dichloro-methane and acetic acid were obtained from POCh (Poland). A CLA isomer mixture, the *cis*-9,*trans*-11 and *trans*-10,*cis*-12 isomers were supplied by Larodan Fine Chemicals AB (Sweden). The purity and details of composition of the applied individual isomers and a mixture of the CLA isomers was examined in our previous study (Czauderna et al., 2003). Sodium selenate (Na₂SeO₄) and fatty acid standards (FA) were provided by Sigma (USA).

HPLC configuration and analytical method

A Waters (Milford, MA, USA) HPLC 625LC system for fractionation of underivatized CLA isomers was used. The system comprised a 515 pump, a 712 WISP auto-sampler and a 2487 dual λ absorbance detector. Two analytical ion-exchange columns loaded with silver ions (250 \times 4.6 mm Chrompac ChromSpher 5 μ m Lipid columns (The Netherlands)) were used in conjunction with a guard column of 10 \times 3 mm (Ag⁺-HPLC system I). The underivatized CLA isomers and other FA-containing conjugated double bonds (CD) in the assayed samples were determined according to Czauderna et al. (2003).

The derivatized CLA isomers and other FA in muscles and fat samples were determined according to Czauderna and Kowalczyk (2002). The HPLC system comprised two Nova Pak C₁₈ columns (4 μ m, 250 \times 4.6 mm I.D., Waters), an alliance separation module (model 2690, Waters) and a Waters 996 photodiode array detector (HPLC system II).

Preparation and hydrolysis of samples

Muscles and fat samples were lyophilized and stored in sealed tubes at -20°C until analysed. Muscle and fat samples (20-50 mg) were hydrolysed with 3.5 ml of 2 M NaOH at 80-85°C for 30 min in sealed tubes. The hydrolysates were acidified with 4 M HCl and then free FA were extracted with dichloromethane (Czauderna and Kowalczyk, 2002). The lower organic layer was dried with Na₂SO₄ and then the organic solvent was removed under a stream of argon. Afterwards the residue was used for derivatization as below or re-dissolved in 1 ml of dichloromethane and then 30 μ l of the resulting solution were injected onto the ion-exchange columns (Ag⁺-HPLC system I) (Czauderna et al., 2003).

Derivatization procedure

To a residue in a reacti-vial, 0.5 ml of dibromacetophenone (48 g/L in acetone) and 60 μ l of triethylamine were added. Next, all FA were derivatized and prepared for HPLC analyses according to Czauderna et al. (2002). The derivatized samples were injected onto C₁₈-Nova Pak columns (HPLC system II) (Czauderna and Kowalczyk, 2002).

Statistical analysis

Statistical analyses of the effects of the CLA isomers or Se were conducted using the nonparametric Mann-Whitney U test for comparing independent experimental groups, while statistical analyses of the simultaneous Se and CLA isomer treatments (Groups 3_{+Se}-6_{+Se}) were performed applying two-factorial analysis for comparison with the independent control group 1. The Statistica (version 6) and Excel 2000 programs were used for the statistical analyses. Differences were considered statistically significant at $P < 0.05$ or at $P < 0.01$.

RESULTS AND DISCUSSION

The effects of experimental diets on body mass gain and feed intake

As can be seen from Table 2, feeding the CLA isomer mixture (Groups 3 and 6), the *trans*-10,*cis*-12 isomer (Group 5), or only Se (Group 2_{+Se}) to rats tended to decrease body weight gain compared with control rats (Group 1). Supplementation of the diet enriched only in the *trans*-10,*cis*-12 isomer most efficiently decreased body weight gain and feed conversion efficiency. These results are supported by several other studies, which have indicated that the *trans*-10,*cis*-12 isomer, due to its geometric and positional structure, is the most potent CLA isomer in term of antiobesity activity, thus, the one which most efficiently reduces feed intake, body weight gain, as well as increases energy expenditure. In contrast, when the *cis*-9,*trans*-11 isomer was added to the rat diet (Group 4) no change in body weight gain or feed conversion efficiency was found, whereas addition of this isomer to the diet enriched in Se (Group 4_{+Se}) reduced the negative effect of Se on body weight gain of rats (Group 2_{+Se}). A similar phenomenon was observed when the CLA isomer mixture was present in the diets enriched in Se (groups 3_{+Se} and 6_{+Se}), moreover, the higher CLA dose (2%) resulted in the smallest body weight gain depression in rats fed Se (Group 6_{+Se}). On the basis of the obtained results, it can be concluded that the interaction between Se and the CLA isomer mixture or the *cis*-9,*trans*-11 isomer reduced the negative effect of the Se-supplemented diet on rat

TABLE 2

Feed intake, body mass gain, feed conversion efficiency in rats fed diet with CLA mixtures (1 and 2%) or individual isomers (1%) (i.e. *c9,t11* or *t10c12*) without Se or with Se (_{+Se}) diets (FCE: g body mass gain/g feed intake)¹

Group	Feed intake for periods, days ²			Total feed intake, g ³	Body mass gain, g	FCE g/g
	1-7	8-14	15-21			
1	125.8 ^{ABCDab}	114.7	100.0	435.3 ^{abc}	59.4 ^a	0.1365
2 _{+Se}	124.3 ^c	116.2 ^b	96.3	432.8	52.8 ^b	0.1220
3	118.2 ^{Ad}	112.0 ^a	97.6	426.5 ^a	54.8	0.1285
4	125.0 ^d	122.0 ^a	98.1	439.5 ^d	59.7	0.1359
5	116.6 ^B	114.1	94.6	420.2	54.1	0.1287
6	116.4 ^C	108.5	96.7	412.9 ^{bd}	56.8	0.1376
3 _{+Se}	118.0 ^a	114.7	100.0	425.4 ^d	56.1	0.1319
4 _{+Se}	123.5	115.7	97.5	430.4	55.5 ^a	0.1289
5 _{+Se}	120.2 ^b	116.1	99.8	435.8	62.2 ^b	0.1427
6 _{+Se}	113.4 ^{Dc}	109.8 ^b	95.6	414.6 ^{cd}	58.4	0.1409

¹ mean values in columns having the same superscripts are significantly different at ^{A,B}P < 0.01 or at ^{a,b}P < 0.05 level

² feed intake in 22-28 days was not included as it did not differ among a group

³ during the whole experimental period (28 days)

growth performance. It was found that the *trans*-10,*cis*-12 isomer in combination with Se (Group 5_{+Se}) produced the highest body weight gain (P < 0.05) as well as the best feed conversion efficiency. Therefore, it seems reasonable to assume that the interaction between Se and the *trans*-10,*cis*-12 isomer or its metabolites resulted in the most efficient elevation of body weight accretion in rats, consequently, most effectively decreased energy expenditure. Rats fed only Se-supplemented diets (Group 2_{+Se}) showed the most distinctly reduced body weight gain, therefore, Se in the diet is responsible for the lowest feed conversion efficiency.

Supplementation of the diets with the *trans*-10,*cis*-12 isomer (Group 5) significantly decreased (P < 0.01) feed intake during the first week of the experiment (Table 2), although no significant decrease was observed after longer experimental times (2-4 weeks). The obtained results are consistent with the above-mentioned data that the *trans*-10,*cis*-12 isomer decreased the body weight gain of rats. Also, feed intake over the period of the first and second week of the study was significantly (P < 0.01 and P < 0.05, respectively) influenced by the inclusion of the CLA isomer mixture into the diet at the lower level (1%), but from week 3 to 4 there was only a tendency towards lower feed intake; however, total feed intake significantly decreased (P < 0.05). Rats fed the diet containing 2% CLA (Group 6) showed a stronger effect on reducing feed intake than those receiving 1% of CLA. However, a lower body weight gain was found in rats receiving 1% of the CLA isomer mixture (Group 3) than animals fed the mixture at 2% in the diet (i.e. in

the diet containing a higher energy level). In the presence of Se, the CLA isomer mixture (1 or 2% in the diet) induced a similar effect on feed intake, however, only the group receiving 2% CLA and Se (Group 6_{+Se}) most efficiently decreased ($P < 0.05$) total feed intake. In contrast, feeding the *cis*-9,*trans*-11 isomer significantly increased ($P < 0.05$) total feed intake, although it had a minute effect on the rats' body weight gain and feed conversion efficiency. The opposite phenomenon was observed in the presence of Se in the diet (Group 4_{+Se}), because this dietary additive resulted in lower feed intake and body gain, therefore, decreased the feed conversion efficiency.

Effects of experimental diets on rat organ weight

The supplemented diets had substantial effects on heart, pancreas, spleen, and liver weight, whereas none of the diets had any effect on kidney or brain weight (Table 3). Heart weight was significantly increased ($P < 0.01$ or $P < 0.05$) by all experimental diets and particularly ($P < 0.01$) by the addition of the *cis*-9,*trans*-11 isomer or 2% CLA to Se-unsupplemented and -supplemented diets (Groups 4, 6 and 4_{+Se}, 6_{+Se}). The increased heart weight associated with the conjugated isomers, regardless of the presence of Se in the diets, is probably due to increased lipoprotein or protein content in this organ (West et al., 1998). The action of the *trans*-10,*cis*-12 isomer in the Se-supplemented diet on the increase of pancreas and liver weight ($P < 0.01$) is likely due to lipid accumulation in these organs. In the work of West et al. (1998), it was argued that liver lipid accumulation could be explained by dietary manipulations, including body mass loss, as well as modifications of protein and fat composition. As the CLA isomers, like Se, modulated immune function, probably through cytokines (West et al., 1998), spleen weight

TABLE 3

Average body mass, and organ mass of rats after 4 weeks of the experiment¹

Group	Rat mass	Heart	Pancreas	Spleen	Liver	Kidneys	Brain
1	241	0.775 ^{ABCdabc}	0.880 ^{ABCD}	0.464 ^{Aa}	9.09 ^{AB}	1.79	1.76
2 _{+Se}	238	0.833 ^{ad}	0.943	0.486 ^b	9.47	1.77	1.75
3	239	0.856 ^b	0.940	0.490	9.33	1.77	1.75
4	245	0.897 ^A	0.889	0.479	9.36	1.73	1.72
5	238	0.844 ^c	1.006 ^B	0.496	9.48	1.81	1.74
6	240	0.911 ^B	0.897	0.520	9.44	1.75	1.75
3 _{+Se}	238	0.859 ^C	1.005 ^A	0.471	9.15	1.72	1.76
4 _{+Se}	240	0.928 ^{Dd}	0.940	0.512 ^a	9.52	1.85	1.75
5 _{+Se}	246	0.876 ^E	1.038 ^C	0.515 ^{Ab}	10.09 ^A	1.79	1.74
6 _{+Se}	241	0.921 ^F	1.023 ^D	0.518	9.80 ^B	1.73	1.78

¹ means in columns with the same letter are significantly different at ^{A,B} $P < 0.01$ or at ^{a,b} $P < 0.05$ level (derived from fresh organs mass of rats normalized to 100 g of rat)

also increased in rats fed the CLA isomers. High increases of spleen weight were found in rats fed the 2% CLA isomer mixture, regardless of the presence of Se or the *trans*-10,*cis*-12 isomer in the Se-supplemented diet. We suppose that the mixture of CLA isomers or simultaneous dosage of Se and the *cis*-9,*trans*-11 or *trans*-10,*cis*-12 isomer most efficiently stimulated the immune function of rats.

The concentration of conjugated CLA metabolites in rat femoral muscles and perigonadal fat

Detailed investigations using *pre*-column derivatization chromatography have demonstrated that unidentified non-CLA isomers containing conjugated double bonds (CD) in fat and muscles had retention times of 23.2 ± 0.3 , 24.8 ± 0.3 , 38.3 ± 0.4 and 40.3 ± 0.4 min (Czuderna and Kowalczyk, 2002). Therefore, we suggested that these compounds contained more than two double bonds because of their shorter elution times compared with retention times of the CLA isomers. The CLA isomers eluted as six peaks and appeared at around 49–55 min of an HPLC run (Czuderna and Kowalczyk, 2002). The results of earlier studies Banni et al. (1999) and Sébédio et al. (1999) also seem to support our above assumption. Moreover, Gnädig (2002) reported that Wistar rats fed a CLA isomer mixture for one month revealed the presence of conjugated fatty acids: C18:2 (*cis*-6,*cis*-9,*cis*-11 C18:3 and *cis*-6, *trans*-10, *cis*-12 C18:3), C20:3 (*cis*-8,*cis*-11,*trans*-13 C20:3 and *cis*-8,*trans*-12,*cis*-14 C20:3) and C20:4 (*cis*-5,*cis*-8,*cis*-11,*trans*-13 C20:4 and *cis*-5,*cis*-8,*trans*-12,*cis*-14 C20:4). Recent results on this subject also reinforce the finding that CLA isomers can be metabolized *in vivo* into long-chain PUFA using the same pathway as linoleic acid (Gnädig, 2002; Bawa, 2003). Therefore, in the current study we could successfully investigate the effect of experimental diets on the content of the sum of CD in muscles and fat of rats (Table 4). The study showed that the level of CD in muscles and fat of control rats (Group 1) was very low, while a small insignificant increase was found in the muscle of rats fed Se (Group 2_{+Se}). As expected, the CLA isomer mixture or individual isomers with or without Se treatments resulted in a significant increase ($P < 0.01$) in the concentration of CD in muscles and fat compared with the control rats (Group 1) and rats fed Se (Group 2_{+Se}). Interestingly, this study is unique in that the animal diets enriched in Se regardless of the presence of the CLA isomers had a major impact on the CD and CLA isomer content in muscles and fat. The addition of Se to the diets enriched in CLA isomers resulted in elevated concentrations of CD and a mixture of CLA isomers in muscles, moreover, the highest significant increase ($P < 0.01$) in CD was found in rats fed diets containing the *cis*-9,*trans*-11 isomer and Se (Group 4_{+Se}). A possible explanation may be that Se supplementation to the experimental diets stimulated the yield of the conversion of CLA isomers into metabolites containing conjugated double bonds (CD) using the same pathway as linoleic acid.

TABLE 4
Total CLA isomers content, individual CLA isomers (c9,t11 and t10,c12) and other non-CLA isomers containing conjugated double bonds (CD) concentrations (mg/g) of femoral muscles and perigonadal fat of rats fed experimental diets²

Group	Muscle ³		Perigonadal fat ³		Muscle ⁴		Perigonadal fat ⁴	
	CLA	CD	CLA	CD ⁵	c9,t11 isomer	t10,c12 isomer	c9,t11 isomer	t10,c12 isomer
1	0.20 ± 0.14	0.14 ± 0.09	0.30 ± 0.12	0.26 ± 0.12	0.055 ± 0.045	0.060 ± 0.075	0.21 ± 0.15	0.13 ± 0.14
2 _{-Se}	0.22 ± 0.21	0.20 ± 0.11 ^{ABCD}	0.20 ± 0.04	0.28 ± 0.23 ^{ABCa}	0.10 ± 0.14	0.051 ± 0.039	0.11 ± 0.48	0.51 ± 0.24
3	4.9 ± 6.4 ^{Aa}	0.46 ± 0.19	16.9 ± 5.7	0.91 ± 0.34	1.7 ± 0.3 ^{Aa}	1.3 ± 0.3(0.76) ⁶	10.5 ± 3.8 ^a	7.3 ± 3.0(0.70) ⁶
4	6.2 ± 1.8	0.40 ± 0.13 ^E	14.5 ± 1.5	0.49 ± 0.09	4.5 ± 1.5	0.20 ± 0.15	15.5 ± 7.2	0.9 ± 1.1
5	5.8 ± 2.3 ^b	0.54 ± 0.29 ^a	18.9 ± 6.8	1.4 ± 1.0	0.13 ± 0.07 ^C	4.0 ± 1.9	0.52 ± 0.20	22.2 ± 9.2
6	9.3 ± 1.5 ^A	0.60 ± 0.14	24.1 ± 9.1	2.0 ± 0.3	3.4 ± 0.8 ^{Aa}	2.6 ± 0.5(0.75) ⁶	13.0 ± 4.6	10.6 ± 3.9(0.82) ⁶
3 _{+Se}	7.4 ± 2.4 ^b	0.57 ± 0.11 ^A	14.0 ± 4.0	0.77 ± 0.17 ^A	3.0 ± 1.0 ^B	2.2 ± 0.7(0.71) ⁶	6.0 ± 2.4 ^a	4.8 ± 1.9(0.80) ⁶
4 _{+Se}	6.9 ± 2.9	1.3 ± 0.5 ^{BE}	15.2 ± 6.7	0.70 ± 0.38 ^a	5.3 ± 2.6	0.17 ± 0.7	13.4 ± 10.1	1.0 ± 4
5 _{+Se}	8.6 ± 2.7 ^b	1.2 ± 0.4 ^{Ca}	13.9 ± 2.5	0.92 ± 0.16 ^B	0.53 ± 0.27 ^C	6.0 ± 2.3	1.1 ± 0.7	18.3 ± 10.9
6 _{+Se}	17.4 ± 8.3	1.1 ± 0.7 ^D	23.7 ± 8.2	1.0 ± 0.3 ^C	6.3 ± 3.5 ^a	5.1 ± 2.5(0.81) ⁶	12.7 ± 6.5	9.9 ± 4.8(0.78) ⁶

¹ fatty acid content (mg) was calculated from lyophilized samples (g) (i.e. fatty acid mg/g of muscle or perigonadal fat); mean ± SD
² means in columns with the same letter are significantly different at ^{A,B}P < 0.01 or at ^{a,b}P < 0.05 level
³ all diets supplemented with CLA isomers or CLA isomer supplemented diets enriched in Se highly significantly increased CLA or CD content in all assayed samples compared with the control rats (Group 1) or the Se supplemented diet (Group 2_{-Se}), respectively (the significant difference at the P < 0.01 level)
⁴ all diets supplemented with CLA isomers or CLA isomer supplemented diets enriched in Se highly significantly increased CLA isomers content in all assayed samples compared with the control rats (Group 1) or the Se supplemented diet (Group 2_{-Se}), respectively (the significant difference at the P < 0.01 level)
⁵ non-CLA isomers containing conjugated double bonds
⁶ concentration ratio of the *cis*-9,*trans*-11 isomer (c9,t11) to the *trans*-10,*cis*-12 isomer (t10,c12) in assayed samples

In contrast, feeding the CLA isomer mixture- or the *trans*-10,*cis*-12 isomer-enriched diets containing Se resulted in decreasing deposition of CD and the sum of all CLA isomers in rat fat. Feeding the *cis*-9,*trans*-11 isomer and Se (Group 4_{+Se}) to rats also resulted in an increase in the concentrations of CD and the sum of all CLA isomers in rat fat. This confirms our earlier results for rat muscles. The diet enriched with Se and the *cis*-9,*trans*-11 isomer most efficiently stimulated the formation of metabolites containing conjugated double bonds (see Group 4_{+Se} in Table 4). More recently it was shown that the two major CLA isomers: *cis*-9,*trans*-11 and *trans*-10,*cis*-12 are substrates for Δ 5- and Δ 6-desaturase as is linoleic acid (Lor and Herbein, 2003). However, considering the above results, it seems reasonable to suggest that the presence of Se and only the *cis*-9,*trans*-11 isomer in the diet most efficiently elevated the yield of the reaction of the *cis*-9,*trans*-11 isomer with Δ 5- and Δ 6-desaturases, i.e. the rate-limiting enzymes determining the yield of conversion of the CLA isomers to products (CD) possessing conjugated double bonds (Group 4_{+Se}).

The effect of the control diet (Group 1) on the level of CLA isomers was similar to the influence of this diet on the concentration of CD in muscle and fat. The lowest contents of all CLA isomers in rat muscle were found in animals fed the control diet (Group 1), while the Se-supplemented diet (Group 2_{+Se}) tended to slightly increase of sum of CLA isomers and the *cis*-9,*trans*-11 isomer in rat muscle (Table 4). The increase of the dietary level of the CLA mixture induced a significant rise ($P < 0.01$) of the CLA isomer content in muscle. The concentration of total CLA isomers was approximately three times greater than the *cis*-9,*trans*-11 or *trans*-10,*cis*-12 isomer. This may be because the two major CLA isomers: *cis*-9,*trans*-11 and *trans*-10,*cis*-12 isomers can be efficiently converted to other geometric or/and positional isomers of CLA. On the other hand, the total CLA isomer level in perigonadal fat is similar to the sum of the *trans*-10,*cis*-12 and *cis*-9,*trans*-11 isomers. Moreover, regardless of the presence of Se in the diets, the level of individual and total CLA isomers is considerably higher in fat compared with their concentrations in rat muscles. Therefore, we suggest that feeding individual CLA isomers or their mixture resulted in deposition of the CLA isomers in rat fat without noticeable metabolism of the CLA isomers, while the opposite effect was observed in rat muscles, indicating efficient isomerization followed by elongation and catabolism of the dietary CLA isomers.

In our investigations, the ratio of the concentrations (R_{sample}) of the *cis*-9,*trans*-11 isomer to the *trans*-10,*cis*-12 isomer in fat and muscles of rats fed the CLA isomer mixture (Groups 3, 6, 3_{+Se}, 6_{+Se}) decreased compared with the concentration ratio (R_{standard}) of these isomers in the standard mixture of CLA isomers in the rat diets ($R_{\text{sample}} \approx 0.8$ vs $R_{\text{standard}} = 1.0242$; see Tables 1 and 4). Our results are in agreement with the recent study of Martin et al. (2000) who also found that the *trans*-10,*cis*-12 isomer tends to be lower than the *cis*-9,*trans*-11 isomer in rat lipids because the

trans-10,*cis*-12 and *trans*-10,*trans*-12 isomers are preferentially driven through the β -oxidation pathway compared with the *cis*-9,*trans*-11 or *trans*-9,*cis*-11 isomers. An interesting finding of our study was that the response to individual isomers and to the CLA isomer mixture was higher in muscles of rats on the Se-supplemented diets compared with Se-unsupplemented treatments. These results reinforce the hypothesis that Se decreased β -oxidation of the CLA isomers in muscles. Moreover, due to the increase of the CLA isomer (substrate) concentration, the yield of conversion of isomers to long-chain PUFA containing conjugated double bonds (CD) was enhanced. This is in agreement with the results obtained in rat fat showing that Se insignificantly decreased deposition of total CLA isomers and the *trans*-10,*cis*-12 isomer in fat, while the level of the *cis*-9,*trans*-11 isomer was practically unchanged (Table 4) as the major changes in composition of deposited lipids were associated with the *trans*-10,*cis*-12 and *trans*-10,*trans*-12 isomers (Simon et al., 2000; Bell and Kennelly, 2003; Loor and Herbein, 2003).

Effect of the CLA isomers and Se on fatty acid composition

In our study, the fatty acid composition of muscles (Table 5) and fat (Table 6) was affected most by feeding CLA isomers and/or Se. Indeed, feeding both individual isomers and their mixture increased the concentration of all assayed FA in muscles compared with the control rats, though the presence of Se in the CLA-isomer-enriched diets resulted in a greater increase in the concentration of all FA. A significant increase was found in the muscle of rats fed Se and 2% of the CLA isomer mixture ($P < 0.01$) or the *trans*-10,*cis*-12 isomer ($P < 0.05$) (i.e. Groups 6_{+Se} and 5_{+Se}). A similar effect of Se in the diets was observed on the content of stearic acid and total assayed saturated fatty acids (SFA) in rat muscles (Table 5). This finding is consistent with the effect of Se on the CLA isomers in rat muscle (see Table 4) and is probably related to the inhibition of fatty acid β -oxidation in skeletal muscles. Our Se-supplemented diet (Group 2_{+Se}) also evaluated the quantitative significance of changes in the level of fatty acids in muscles (Table 5) and perigonadal fat (Table 6). The effect of supplementing only Se was relatively minor, however consistent. Indeed, in rat muscles and fat, supplementing only Se (VI) slightly reduced the concentration of all assayed FA, while more efficiently lowering the content of C18:0 and SFA (Group 2_{+Se}). Our experiment demonstrated that Se (VI), as a strong oxidant, more efficiently decreased the content of SFA than unsaturated FA. This consideration is supported by the ratios (R) of C18:0 or SFA to unsaturated FA (*cis*-monounsaturated FA and PUFA) because the ratio (R) generally decreased mainly due to the lower level of SFA in muscles (Table 5).

The values of the R ratios summarized in Table 6 varied inconsistently because their values depend upon material changes in the levels of both saturated and unsaturated FA. The diets enriched in individual CLA isomers or their mixture

TABLE 5
Fatty acid (FA) content (mg) and ratios (R) of selected FA in femoral muscles (g) of rats fed diets enriched in the CLA isomers and/or Se¹

Group	ΣFA ² mg/g	C18:0 ³ mg/g	SFA ³ mg/g	R _{C18:0/c9} ⁴	R _{SFA/c9}	R _{C18:0/Σm-c} ⁵	R _{SFA/Σm-c}	R _{C18:0/PUFA}	R _{SFA/PUFA}
1	20 ± 6 ^a	1.4 ± .4	7.1 ± 1.6 ^a	0.313 ^{ABab}	1.671 ^{abcde}	0.275 ^{Aa}	1.466 ^{abc}	0.104 ^{abc}	0.557 ^{ABbb}
2 _{+Se}	19 ± 3 ^{bbc}	1.2 ± .2 ^{Aa}	6.3 ± 1.0 ^{bc}	0.291 ^{Ccd}	1.507 ^{ABCg}	0.291 ^{Bbc}	1.278 ^{ABde}	0.093 ^{ABCd}	0.453 ^{CDfBb}
3	22 ± 2 ^C	1.3 ± .1	6.1 ± 0.8 ^d	0.441 ^A	2.147 ^a	0.393 ^A	1.896 ^a	0.080 ^D	0.386 ^G
4	24 ± 5	1.1 ± .2	6.2 ± 1.0	0.321	1.722	0.290	1.556	0.068	0.367
5	25 ± 8	1.4 ± .4	7.3 ± 1.8	0.413 ^a	2.220 ^b	0.342	1.836 ^b	0.076	0.418
6	27 ± 3	1.3 ± .1 ^b	6.2 ± 0.7 ^c	0.452 ^B	2.209 ^c	0.375 ^a	1.837	0.062 ^{DE}	0.303 ^G
3 _{+Se}	28 ± 8 ^{Cb}	1.4 ± .4	7.6 ± 2.1 ^d	0.406 ^{Cb}	2.147 ^{UA}	0.339 ^B	1.794 ^{Ac}	0.071 ^{Aa}	0.375 ^{Ca}
4 _{+Se}	27 ± 9	1.3 ± .3	7.3 ± 2.4	0.374	1.980 ^B	0.336 ^b	1.782 ^B	0.073 ^d	0.387 ^D
5 _{+Se}	35 ± 10 ^{Ac}	1.8 ± .4 ^a	9.7 ± 2.7 ^{ac}	0.385 ^c	2.081 ^{cC}	0.321	1.733 ^d	0.071 ^{Bb}	0.383 ^{AE}
6 _{+Se}	41 ± 21 ^{Ba}	1.8 ± .6 ^{Ab}	9.6 ± 4.3 ^{bc}	0.479 ^d	2.002 ^g	0.408 ^c	2.002 ^c	0.064 ^{CcC}	0.319 ^{Bf}

¹ means in columns with the same letter are significantly different at ^{A,B}p < 0.01 or at ^{a,b}p < 0.05 level

² sum of all assayed saturated fatty acids (SFA), MUFA, CLA, CD and PUFA (Czauderna and Kowalezyk, 2002)

³ C18:0 - stearic acid; SFA - sum of C8:0, C10:0, C12:0, C14:0, C16:0, C18:0, C20 and C22:0 (Czauderna and Kowalezyk, 2002)

⁴ c9 - oleic acid (*cis*-9 octadecenoic acid)

⁵ Σm-c - sum of *cis*-monounsaturated FA, i.e. *cis*-vaccenic acid (*cis*-11 octadecenoic acid), oleic acid and petroselinic acid (*cis*-6 octa-decenoic acid)

TABLE 6

Fatty acid concentrations¹ and ratios of selected fatty acids in perigonadal fat from rats fed diet supplemented with CLA isomers and/or Se^{2,3}

Group	ΣFA mg/g	C18:0 Mg/g	SFA mg/g	R _{C18:0/e9}	R _{SFA/e9}	R _{C18:0/Σm-c}	R _{SFA/Σm-c}	R _{C18:0/PUFA}	R _{SFA/PUFA}
1	20 ± 6 ^{3a}	1.4 ± 0.4	7.1 ± 1.6 ^a	0.313 ^{ABab}	1.671 ^{abcde}	0.275 ^{Aa}	1.466 ^{abc}	0.104 ^{abc}	0.557 ^{ABbb}
2 _{+Se}	62 ± 35	2.0 ± 1.3	17 ± 9 ^{abc}	0.117 ^{ABCDabc}	1.055 ^{ABCDabcd}	0.101 ^{ABCDabc}	0.916 ^{ABCDabc}	0.047 ^{abcd}	0.426 ^{ABCD Eab}
3	62 ± 23	1.7 ± 0.5 ^a	11 ± 4	0.196 ^a	1.339 ^a	0.161 ^a	1.101 ^{Garf}	0.037 ^{af}	0.254 ^A
4	43 ± 4	1.2 ± 0.1	21 ± 18	0.164 ^A	2.781 ^B	0.164 ^A	2.780 ^{AH}	0.038 ^b	0.635 ^G
5	58 ± 20	2.0 ± 0.8	13 ± 4	0.229 ^B	1.510 ^C	0.229 ^B	1.510 ^B	0.046	0.302 ^B
6	72 ± 71	1.1 ± 0.2 ^a	8 ± 2	0.215 ^b	1.513 ^c	0.185 ^b	1.300 ^{bf}	0.027 ^{cf}	0.192 ^C
3 _{+Se}	40 ± 10	1.2 ± 0.3	8.8 ± 2.2 ^a	0.200 ^{CE}	1.441 ^{AE}	0.201 ^{CE}	1.441 ^{CEG}	0.039 ^d	0.283 ^D
4 _{+Se}	43 ± 17	1.2 ± 0.4	8.9 ± 3.3 ^b	0.176 ^{Fc}	1.340 ^{be}	0.149 ^{cd}	1.137 ^{Hcd}	0.035 ^e	0.268 ^{EGc}
5 _{+Se}	57 ± 27 ^b	1.6 ± 0.4	11 ± 2 ^d	0.256 ^{DG}	1.801 ^{DF}	0.213 ^{DF}	1.500 ^{DF}	0.043	0.299 ^a
6 _{+Se}	49 ± 13 ^a	2.1 ± 3.6	9 ± 4 ^c	0.528	2.636 ^{Gd}	0.445	2.268 ^e	0.059	0.214 ^b

¹ fatty acid mg/g of perigonadal fat

² abbreviation for fatty acids see Table 5

³ means in columns with the same letter are significantly different at ^(A,B)p < 0.01 or at ^(a,b)p < 0.05 level

generally increased the ratios of C18:0 or SFA to oleic acid and to the sum of *cis*-monounsaturated FA (*cis*-MUFA) in muscles and fat, while decreasing the ratios of C18:0 or SFA to PUFA (i.e. non-conjugated PUFA). Considering the results summarized in Tables 5 and 6, we suggest that individual isomers (probably especially the *trans*-10,*cis*-12 isomer) or a mixture of CLA isomers in the experimental diets caused a reduction in $\Delta 9$ -desaturase capacity, inhibited stearyl-CoA desaturase mRNA expression and fatty acid synthase (Simon et al., 2000; Loor and Herbein, 2003), so the muscle and fat concentrations of both oleic acid and sum the of *cis*-MUFA decreased. The obtained results fully confirmed our above suggestion as higher values of the ratio (R) of C18:0 to oleic acid were observed in muscles (Table 5) and fat (Table 6) of rats fed the *trans*-10,*cis*-12 isomer (Group 5) compared with the *cis*-9,*trans*-11 isomer (Group 4).

The diets supplemented with individual CLA isomers or their mixture had a synergetic effect on PUFA accretion in muscles, while practically not affecting the level of deposited PUFA in perigonadal fat. In our study, CLA isomer treatment generally resulted in a considerable decrease in the concentrations of SFA in fat, whereas minor insignificant reductions were observed in rat muscles.

Feeding Se and individual CLA isomers or their mixture resulted in increasing the sum of both *cis*-MUFA and PUFA in muscles (Table 5) compared with the control rats (Group 1) and/or rats fed the CLA isomers (Groups 3, 4, 5 and 6). Therefore, considering all presented results it seems likely that the diets enriched in Se and individual CLA isomers or their mixture stimulated lipogenesis, desaturation and elongation as well as inhibited FA β -oxidation in muscles. On the other hand, Se in the diets enriched in individual or mixtures of CLA isomers generally resulted in decreasing both the level of total assayed SFA, oleic acid, *cis*-MUFA and PUFA in perigonadal fat (Table 6) compared with the control rats (Group 1) and/or rats fed only the CLA isomers (Groups 3, 4, 5 and 6). Consequently, the changes of ratios (R) are not consistent, however, generally, increased for values of $R_{C18:0/c9}$, $R_{SFA/c9}$, $R_{C18:0/\Sigma m-c}$ and $R_{SFA/\Sigma m-c}$, while decreased for $R_{C18:0/PUFA}$ and $R_{SFA/PUFA}$. Therefore, we could hypothesize that in perigonadal fat, the diets supplemented with Se and individual CLA isomers or their mixture enhanced lipolysis at the expense of FA esterification and reduced the activities of $\Delta 9$ -, $\Delta 6$ - and/or $\Delta 5$ -desaturases.

CONCLUSIONS

The present study demonstrated that the *trans*-10,*cis*-12 and the CLA isomer mixture induced a decrease in body weight gain and food intake, while the *cis*-9,*trans*-11 isomer did not change body mass accretion or food intake. The *cis*-9,*trans*-11 isomer is the most potent CLA isomer possessing anticarcinogenic pro-

perties, while feeding this isomer had no effect on lipid-metabolizing enzymes in any of the organs or tissues of rats. The *cis-9,trans-11* isomer showed a minor role in mediating many of the biochemical effects attributed to the *trans-10,cis-12* isomer. A 12-double bond appears to be a key structure for inhibiting stearyl-CoA desaturase activity, especially when coupled with a 10-double bond, but not with a 9-double bond (Alasnier et al., 2002). Due to the geometric and positional structure, the *trans-10,cis-12* isomer is a more potent antiobesity agent in mammals. This isomer increased FA β -oxidation at the expense of FA esterification, increased hepatic and adipose carnitine palmitoyl-CoA transferase activity, reduced excess body gain, increased energy expenditure, increased feed efficiency and lipolysis (West et al., 1998; Alasnier et al., 2002). Both main isoforms of a mixture of CLA isomers can reduce circulating leptin levels (by decreasing leptin gene expression), therefore, this decrease resulted in reducing the adiposity and body weight of the examined animals (Rahman et al., 2001; Corino et al., 2002).

Based on the above observations, we suggest that simultaneous dietary supplementation of Se and the *trans-10,cis-12* isomer or the CLA isomer mixture had a beneficial effect on accumulation of the *trans-10,cis-12* isomer and the CLA isomer mixture in muscles. We suggest that the interaction between Se and a mixture of CLA isomers or the *trans-10,cis-12* isomer most effectively protected CLA isomers from peroxidation damage and/or catabolism in muscles. The finding that Se and individual CLA isomers or their mixtures fed to animals increased the level of CLA isomers, CD, *cis*-MUFA and PUFA in the muscles of rats is valuable for nutritionists in the context of research to improve the nutritional quality of food for human health.

REFERENCES

- Arteel G.E., Sies H., 2001. The biochemistry of selenium and glutathione system. *Environ. Toxicol. Pharmacol.* 10, 153-158
- Alasnier C., Berdeaux O., Chardigny J.M., S eb edio J.L., 2002. Fatty acid composition and conjugated linoleic acid content of different tissues in rats fed individual conjugated linoleic acid isomers given as triacylglycerols. *J. Nutr. Biochem.* 13, 337-345
- Bauman D.E., Baumgard L.H., Corl B.A., Griinari J.M., 2000. Biosynthesis of conjugated linoleic acid in ruminants. *Proc. Amer. Soc. Anim. Sci.* 1999, 1-15 (online publication)
- Bawa S., 2003. An update on the beneficial roles of conjugated linoleic acid (CLA) in modulating human health: mechanisms of action - a review. *Pol. J. Food Nutr. Sci.* 12, 3-13
- Bell J.A., Kennelly J.J., 2003. Postprandial infusion of conjugated linoleic acids negatively impacts milk synthesis in Holstein cows. *J. Dairy Sci.* 86, 1321-1324
- Corino C., Mourou J., Magni S., Pastorelli G., Rosi F., 2002. Influence of dietary conjugated linoleic acid on growth, meat quality, lipogenesis, plasma leptin and physiological variables of lipid metabolism in rabbits. *J. Anim. Sci.* 80, 1020-1028

- Crespo A.M., Reis M.A., Lanca M.J., 1995. Effect of selenium supplementation on polyunsaturated fatty acids in rats. *Biol. Tr. Elem. Res.* 47, 335-341
- Czauderna M., Kowalczyk J., 2002. HPLC separation of some unsaturated and saturated fatty acids. *Chem. Anal. (Warsaw)* 47, 867-882
- Czauderna M., Kowalczyk J., Wąsowska I., Niedźwiedzka K.M., 2002. A highly efficient method for derivatization of fatty acids for high performance liquid chromatography. *J. Anim. Feed Sci.* 11, 517-526
- Czauderna M., Kowalczyk J., Wąsowska I., Niedźwiedzka K.M., 2003. Determination of conjugated linoleic acid isomers by liquid chromatography and photodiode array detection. *J. Anim. Feed Sci.* 12, 269-382
- DeLany J.P., Blohm F., Truett A.A., Scimeca J.A., West D.B., 1999. Conjugated linoleic acid rapidly reduces body fat content in mice without affecting energy intake. *Amer. J. Physiol.* 276, R1172-R1179
- Evans M.E., Brown J.M., McIntosh M.K., 2002. Isomer-specific effects of conjugated linoleic acid (CLA) on adiposity and lipid metabolism. *J. Nutr. Biochem.* 13, 508-516
- Gnäding S., 2002. Conjugated linoleic acid (CLA): effect of processing on CLA in cheese and the impact of CLA on the arachidonic acid metabolism. PhD Dissertation, Universität Hamburg. INRA, Unité de Nutrition Lipique, Dijon (France)
- Gnäding S., Rickert R., Sébédio J.L., Steinhart H., 2001. Conjugated linoleic acid (CLA): physiological effects and production. *Eur. J. Lipid Sci. Tech.* 103, 56-61
- Jahreis G., Fritsche J., Steinhart H., 1997. Conjugated linoleic acid in milk fat: high variation depending on production system. *Nutr. Res.* 17, 1479-1484
- Lawson R.E., Moss A.R., Givens D.I., 2001. The role of dairy products in supplying conjugated linoleic acid to man's diet: a review. *Nutr. Res. Rev.* 14, 153-172
- Loor J.J., Herbein J.H., 2003. Reduced fatty acid synthesis and desaturation due to exogenous *trans*10,*cis*12-CLA in cows fed oleic or linoleic oil. *J. Dairy Sci.* 86, 1354-1369
- Martin J.C., Gregoire S., Siess M.H., Genty M., Chardigny J.M., Berdeaux O., Juaneda P., Sébédio J.L., 2000. Effects of conjugated linoleic acid isomers on lipid metabolizing enzymes in male rats. *Lipids* 35, 91-98
- Pastuszewska B., Ochtabińska A., Morawski A., 2000. A note on the nutritional adequacy of stock diets for laboratory rats and mice. *J. Anim. Feed Sci.* 9, 533-542
- Rahman S.M., Wang Y.M., Yotsumoto H., Cha J.Y., Han S.Y., Inoue S., Yanagita T., 2001. Effects of conjugated linoleic acid on serum leptin concentration, body-fat accumulation, and β -oxidation of fatty acid in OLETF rats. *Nutrition* 17, 385-390
- Simon O., Männer K., Schäfer K., Sagredos A., Eder K., 2000. Effects of conjugated linoleic acids on protein to fat proportions, fatty acids, and plasma lipids in broilers. *Eur. J. Lipid Sci. Tech.* 102, 402-410
- Sébédio J.L., Gnäding S., Chardigny J.M., 1999. Recent advances in conjugated linoleic acid research. *Curr. Opin. Clin. Nutr. Metabol. Care* 2, 499-506
- Szymczyk B., Pisulewski P., Szczurek W., Hanczakowski P., 2000. The effects of feeding conjugated linoleic acid (CLA) on rat growth performance, serum lipoproteins and subsequent lipid composition of selected rat tissues. *J. Sci. Food Agr.* 80, 1553-1558
- Tsuboyama-Kasaoka N., Takahashi M., Tanemura K., Kim H.J., Tange T., Okuyama H., Kasai M., Ikemoto S., Ezaki O., 2000. Conjugated linoleic acid supplementation reduces adipose tissue by apoptosis and develops lipodystrophy in mice. *Diabetes* 49, 1534-1542
- West D.B., DeLany J.P., Camet P.M., Blohm F., Truett A.A., Scimeca J., 1998. Effects of conjugated linoleic acid on body fat and energy metabolism in the mouse. *Amer. J. Physiol.* 275, R667-R672

- Wolfram S., 1999. Absorption and metabolism of selenium: differences between inorganic and organic sources. In: T.P. Lyons, K.A. Jacques (Editors). *Biotechnology in the Feed Industry*. Nottingham University Press, pp. 547–566
- Zagrodzki P., Bik D., Fitak B.A., Suchocki P., Niemczuk K., 2000. Selenoenzymes in animal tissues after supplementation with selol. *Bull. Vet. Inst. Pulawy* 44, 215-220
- Zagrodzki P., Nicol F., Arthur J.R., Słowiaczek M., 2001. Selenoproteins in human thyroid tissues. *Biofactors* 14, 223-227

STRESZCZENIE

Wpływ selenu i izomerów sprzężonego kwasu linolowego (CLA) w diecie na zawartość kwasów tłuszczowych i izomerów CLA w tkankach szczurów oraz na ich wzrost

Badano wpływ izomerów sprzężonego kwasu linolowego (CLA) lub/i selenu (w postaci Na_2SeO_4) na wzrost szczurów i poziom wybranych kwasów tłuszczowych (KT) w mięśniu udowym oraz tłuszczu okołogonadowym. Podawanie zwierzętom mieszaniny izomerów CLA, izomeru *trans*-10,*cis*-12 lub selenu (Se) powoduje zmniejszenie przyrostu masy ciała szczurów w porównaniu z grupą kontrolną. Przy jednoczesnym dodatku izomeru *trans*-10,*cis*-12 i Se do dawki przyrosty masy ciała zwierząt były najwyższe, a wykorzystanie paszy najlepsze. Dodatek izomerów CLA i/lub Se do diety nie miał wpływu na masę mózgu i nerek, natomiast zwiększał masę trzustki, wątroby i śledziony. Izomery CLA dodane do dawki istotnie zwiększyły w badanych tkankach stężenie izomerów CLA i innych kwasów tłuszczowych zawierających sprzężone podwójne wiązania (CD). Jednoczesne podanie Se i izomerów CLA stymuluje gromadzenie się izomerów CLA i CD w mięśniach, natomiast podawanie jedynie izomeru *cis*-9,*trans*-11 i Se stymulowało akumulację izomerów CLA i CD w tłuszczu. Badania dowiodły, że izomer *trans*-10,*cis*-12 wydajniej ulega β -oksydacji w porównaniu z izomerem *cis*-9,*trans*-11. Izomery CLA bez względu na obecność Se w dawce zwiększają zawartość KT w mięśniach, natomiast zmniejszają zawartość nasyconych KT w tłuszczu. Jednoczesne podawanie Se i izomerów CLA zwiększa stężenie *cis*-mononienasyconych KT (*cis*-MNKT), CD i wielonienasyconych KT (WNKT) w mięśniu, natomiast zmniejsza zawartość nasyconych KT, *cis*-MNKT i WNKT w tłuszczu. Jednoczesne podanie Se i mieszaniny CLA izomerów lub izomeru *trans*-10,*cis*-12 najefektywniej chroni izomery CLA przed oksydacyjnym uszkodzeniem oraz hamuje ich katabolizm.