

A note on chromosome abnormalities in early turkey embryos

K. Jaszczak¹, R. Parada and M. Sacharczuk

*Department of Molecular Cytogenetics,
Institute of Genetics and Animal Breeding,
Polish Academy of Sciences
Jastrzębiec, 05-552 Wólka Kosowska, Poland*

(Received 10 April 2003; accepted 28 October 2003)

ABSTRACT

Two commercial turkey lines: medium-heavy weight (MHW) and heavy-weight (HW) were used for a karyotype analysis and study of the frequency of chromosome abnormalities in early embryos (24 h). Among the total of 378 embryos, 40 (10.5%) had an abnormal chromosome complement consisting of 14 haploids, 20 haploid-diploid chimeras, 2 triploids and 4 diploid-tetraploid mosaics. A significantly greater frequency of heteroploid embryos was recovered from the MHW (15.4%) than from the HW line (8.0%). The discussion points to the negative effect of chromosome abnormalities on hatchability.

KEY WORDS: embryos, chromosome abnormalities, turkey

INTRODUCTION

An analysis of the chromosome complements of early chicken embryos has revealed that in domestic fowl various types of chromosome abnormalities can occur, including haploidy, triploidy, aneuploidy (Bloom, 1972; Miller et al., 1976). The chromosome abnormalities are caused principally by meiotic nondisjunction, polar body suppression during oogenesis, polyspermy and errors during early cleavage (Fechheimer, 1981). The level of spontaneous chromosome aberrations in chicken varies depending on line, stock, breed and direction of selection (Reddy and Siegel, 1977; Snyder et al., 1979; Jaszczak et al., 1992). Chromosome abnormalities in avian embryos can be responsible for their high mortality and hatching

¹ Corresponding author: e-mail: k.jaszczak@ighz.pl

losses (Thorne et al., 1991; Jaszczak and Jaszczak, 1993). Elimination from the breeding stock of families or individuals producing a high proportion of chromosome abnormalities among the embryos may be of considerable practical importance.

A majority of karyotype studies performed on poultry were related to chicken and sporadically quail or geese (de la Sena et al., 1992; Jaszczak et al., 1999). Cytogenetic investigations of turkey embryos were not conducted, despite the fact that this species is very important for poultry meat production. The aim of the present studies was to estimate the frequency of chromosome abnormalities in early embryos of two turkey commercial lines.

MATERIAL AND METHODS

The cytogenetic study was carried out on embryos from two commercial lines of turkeys: Nicholas 300 (medium-heavy weight MHW) and Nicholas 700 (heavy-weight HW). Samples were collected over a period of one season - from April to July. Eggs were collected and stored at 12°C for two weeks and then placed in an incubator at 38°C. After 24 h incubation the eggs were opened and examined macroscopically. The blastodisks from fertile eggs were removed and each was turned into a cell suspension in 2 ml of culture medium containing colchicine. Further processing included short culture (2 h), hypotonic, fixation and slide preparation, according to the method described by Zartman and Jaszczak (1980). Five to 10 metaphase spreads were analysed by scoring the presence of the 9 largest pairs of autosomes and sex chromosomes ZZ and ZW. When karyotype abnormalities were detected 20 to 30 metaphases were analysed (if available). The karyotype analysis was carried out under a light microscope with CCD camera connected to a computer supplied with a Multiscan analytic programme.

Differences between the two turkey lines were estimated by the chi-square test.

RESULTS AND DISCUSSION

Results of the cytogenetic survey of early embryos (24 h) from two lines of turkeys are given in Table 1. Among 378 turkey embryos 40 (10.5%) were composed of cells with abnormal chromosome complements.

Among 40 embryos with an abnormal karyotype 34 contained haploid cells, either alone or in conjunction with diploid cells. The remaining aberrations included 4 diploid/tetraploid mosaics and 2 triploids. A significantly greater frequency of chromosomal abnormalities was recovered from the MHW (15.4%) than the HW line (8.0%). The frequency of pure haploidy (1A) was specially high in line

MHW, while the frequency of other heteroploidy types were relatively low in both lines. The sex chromosome complements of aberrant turkey embryos are showed in Table 2. In haploid embryos and in haploid cells from the haploid/diploid chimeras only the Z sex chromosome was observed. Chromosome W was not found. This suggests that, similarly as in chicken, haploid cell lines in embryos, either pure haploidy (1A) or haploid/diploid chimerism, are of androgenetic origin (Zartman and Smith, 1975; Fechheimer and Jaap, 1978). The occurrence of two triploid turkey embryos, with only the ZZZ sex chromosomes, might have resulted from diandry or from digyny, originating from a suppression of meiosis II during oogenesis (Fechheimer, 1981; Thorne and Sheldon, 1991; de la Sena et al., 1992). In the 4 diploid/tetraploid turkey embryos the sex chromosome complement of the diploid cells was doubled in the tetraploid cells, what indicates that, similarly as in chicken, tetraploid cells were produced by endoreduplication or suppression of cytokinesis during early cleavage mitosis (Miller et al., 1971). In C- banded diploid/tetraploid quail embryos, C-band variants of chromosome 4 and chromosome Z in diploid cells were also doubled in tetraploid cells (de la Sena et al., 1991).

TABLE 1

Number and percentages (in parentheses) of heteroploid embryos

Abnormality	Line		Total
	medium-heavy weight	heavy weight	
	MHW	HW	
Haploidy (1A)	10 (7.7)	4 (1.6)	14 (3.7)
Haploid/diploid chimerism (1A/2A)	8 (6.1)	12 (4.8)	20 (5.3)
Triploidy (3A)	-	2 (0.8)	2 (0.5)
Diploid/tetraploid mosaicism (2A/4A)	2 (1.5)	2 (0.8)	4 (1.0)
Total heteroploid embryos	20 (15.4) ¹	20 (8.0) ¹	40 (10.5)
Total number of embryos	130	248	378

¹ P ≤ 0.043

TABLE 2

Sex chromosome complement of turkey embryos with chromosome abnormalities

Abnormality	MHW line		HW line	
	number of embryos	sex chromosome complement	number of embryos	sex chromosome complement
	Haploidy (1A)	10	Z	4
Haploid/diploid chimerism (1A/2A)	5	Z/ZZ	8	Z/ZZ
	3	Z/ZW	4	Z/ZW
Triploidy (3A)	-	-	2	ZZZ
Diploid/tetraploid mosaicism (2A/4A)	-	-	1	ZZ/ZZZZ
	2	ZW/ZZWW	1	ZW/ZZWW

In the literature available there is no data concerning chromosome abnormalities in turkey embryos. A comparison of the frequency of chromosome aberrations observed in the present study, with that reported for other bird species, indicates that in turkeys this is a frequent phenomenon (Reddy and Siegel, 1977; Wolowodiuk et al., 1995; Jaszczak et al., 1999). A significantly greater frequency of chromosome abnormalities in the MHW than HW line can result from genetic differences between the components used for creating commercial turkey lines. Individual lines or strains of chicken are characterized by a large variation in the overall rate of chromosome aberrations in embryos (Thorne and Sheldon, 1991; Jaszczak et al., 1993). For that reason it is necessary to conduct further cytogenetic studies of chromosome abnormalities in the parent stock used for creating commercial lines. The results obtained indicate the value of a cytogenetic survey of turkeys aiming at an elimination of the carriers of chromosome aberrations, as those may be the reason of a lower hatchability of fertile eggs.

ACKNOWLEDGEMENTS

The authors thank Zofia Piechal for technical assistance.

REFERENCES

- Bloom S.E., 1972. Chromosome abnormalities in chicken (*Gallus domesticus*) embryos: types, frequencies and phenotypic effects. *Chromosoma* 37, 309-326
- de la Sena C.A., Fechheimer N.S., Nestor K.E., 1991. Variability of C-banding patterns in the Japanese quail. *Genome* 34, 993-997
- de la Sena C.A., Fechheimer N.S., Nestor K.E., 1992. Evidence for genetic etiology of heteroploidy in embryos of the Japanese quail (*Coturnix coturnix japonica*). *Cytogenet. Cell Genet.* 60, 140-145
- Fechheimer N.S., 1981. Origins of heteroploidy in chicken embryos. *Poultry Sci.* 60, 1365-1371
- Fechheimer N.S., Jaap R.G., 1978. The parental source of heteroploidy in chick embryos determined with chromosomally marked gametes. *J. Reprod. Fertil.* 52, 141-146
- Jaszczak K., Cywa-Benko K., Wężyk S., 1992. Chromosome abnormalities in early embryos from chickens maintaining in preservable flocks. *Rocz. Nauk. Zoot., Monogr. Rozpr.* 31, 285-294
- Jaszczak K., Jaszczak J., 1993. Chromosomal abnormalities in early embryos and commercial chicks and hatchability in reproduction flocks of layer and broiler hens. *Genet. Pol.* 34, 287-293
- Jaszczak K., Rabsztyn A., Jaszczak J., Kapkowska E., 1999. A karyotypic survey of early goose embryos. *Anim. Sci. Pap. Rep.* 17, 67-72
- Miller R.C., Fechheimer N.S., Jaap R.G., 1971. Chromosome abnormalities in 16- to 18-hour chick embryos. *Cytogenetics* 10, 121-136
- Miller R.C., Fechheimer N.S., Jaap R.G., 1976. Distribution of karyotype abnormalities in chick embryo sibships. *Biol. Reprod.* 14, 549-560

- Reddy P.R.K., Siegel P.B., 1977. Chromosomal abnormalities in chickens selected for high and low body weight. *J. Hered.* 68, 253-256
- Snyder M.D., Fechheimer N.S., Jaap R.G., 1979. Incidence and origin of heteroploidy, especially haploidy, in chick embryos from intraline and interline matings. *Cytogenet. Cell Genet.* 14, 63-75
- Thorne M.H., Collins R.K., Sheldon B.L., 1991. Chromosome analysis of early embryonic mortality in layer and broiler chicken. *Brit. Poultry Sci.* 32, 711-722
- Thorne M.H., Sheldon B.J., 1991. Cytological evidence of maternal meiotic errors in a line of chickens with a high incidence of triploidy. *Cytogenet. Cell Genet.* 57, 206-210
- Wolodiuk D., Fechheimer N.S., Nestor K.E., Bacon W.L., 1985. Chromosome abnormalities in embryos from lines of Japanese quail divergently selected for body weight. *Genet. Sel. Evol.* 17, 183-190
- Zartman D.L., Jaszczak K., 1980. A procedure for chromosome preparations from early chick embryos. *Pr. Mat. Zoot.* 23, 115-119
- Zartman D.L., Smith A.L., 1975. Triploidy and haploid-triploid mosaicism among chick embryos (*Gallus domesticus*). *Cytogenet. Cell Genet.* 15, 138-145

STRESZCZENIE

Nieprawidłowości chromosomowe we wczesnych zarodkach indyków

Badano kariotypy wczesnych zarodków (24 godz.) z dwóch komercyjnych linii indyków: średnio ciężkiej (Nicholas 300) i ciężkiej (Nicholas 700). Spośród przeanalizowanych łącznie 378 zarodków nieprawidłowości chromosomowe stwierdzono w 40 przypadkach (10,5%); najczęściej występował chimeryzm haploidalno/diploidalny (20) i haploidia (14). W badanym materiale były poza tym dwa zarodki triploidalne i dwa z mozaicyzmem diploidalno/tetraploidalnym. Istotnie większa częstotliwość heteroploidalnych zarodków wystąpiła w linii średnio ciężkiej (15,4%) niż w linii ciężkiej (8,0%). W dyskusji wskazano na negatywny efekt nieprawidłowości chromosomowych w reprodukcji.